

Combined Effects of Succinylcholine and Calcium on Membrane-bound Acetylcholinesterase Activity

Masaki WAKAMATSU, Hiroyuki SHIMONAKA, Michio YAMAMOTO,
Kiyoshi KAWAI* and Yoshinori NOZAWA *

The effects of succinylcholine and calcium (Ca^{2+}), alone and together, on membrane-bound acetylcholinesterase ("true-type" cholinesterase) were examined using human erythrocyte ghosts to elucidate the combined pharmacological activity of succinylcholine and calcium in in vivo system. Succinylcholine inhibited the acetylcholinesterase by a mixed style. Calcium alone exhibited an inhibitory effect on the enzyme, but a biphasic effect together with succinylcholine : marked restoration of the enzyme activity at calcium concentrations lower than 6 mM and depression at its higher concentrations. It is suggested that calcium induces a conformational change of the enzyme protein leading to the altered binding capacity of succinylcholine. In anesthetic practice, therefore, the use of calcium may not be indicated for the treatment of SCh phase II block. (Key words: succinylcholine, calcium, acetylcholinesterase, human erythrocyte membrane)

(Wakamatsu M et al.: Combined effects of succinylcholine and calcium on membrane-bound acetylcholinesterase activity. *J Anesth* 1:15-21, 1987)

Calcium has been shown to affect the neuromuscular blocking action of succinylcholine (SCh). Mayrhofer¹ and Irwin et al.² have reported that calcium decreases the duration of apnea induced by SCh in humans and dogs, respectively while Badola et al.³ have indicated the calcium-induced depression of twitch tension during SCh desensitization block. According to Shrivastava et al.⁴, recently, calcium prevented the SCh-induced myalgia. But few attempts have been made to elucidate interactions between calcium and SCh at the molecular level. Acetylcholinesterase (E.C.3.1.1.7.; AChE), which hydrolyzes and inactivates acetylcholine, plays an important role in the neuromuscular trans-

mission. Several muscle relaxants and metal ions are thought to bind to allosteric sites on AChE and to influence its catalytic properties. Lanks and Sklar⁵ have shown that SCh inhibits the activity of erythrocyte AChE in a non-competitive manner. Calcium has also been known as a modulator of AChE, and it usually activates the enzyme in solubilized preparations⁶. However, there has been no in vitro study of a combination of SCh and calcium, which may serve the pharmacological and physiological evaluation of neuromuscular transmission. It appears that membrane-bound AChE of human erythrocyte can substitute for the detergent-soluble tissue enzyme for in vitro study, though the biological function and significance of the erythrocyte AChE are unknown⁷.

In the present study, we investigated the effects of SCh and calcium, alone and together, on the activity of membrane-bound AChE in human erythrocyte ghost membranes to elucidate the pharmacological interactions between

*Department of Anesthesiology and *Biochemistry, Gifu University School of Medicine, Gifu, Japan*

Address reprint requests to Dr. Wakamatsu: Department of Anesthesiology, Gifu University School of Medicine, 40 Tsukasa-machi, Gifu, 500 Japan

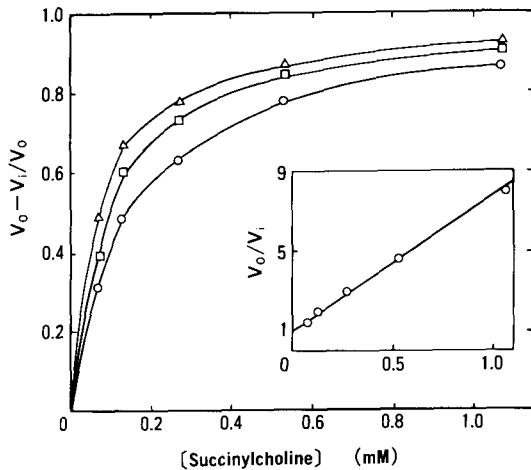


Fig. 1. Effect of succinylcholine on hydrolysis of acetylthiocholine. Inset shows inhibition plot of acetylcholinesterase with succinylcholine at 0.30 mM substrate. Half-maximal inactivation was observed at 0.15 mM succinylcholine. V_i and V_0 represent the initial velocity with and without succinylcholine, respectively. Substrate concentrations were (Δ)0.055 mM, (\square)0.11 mM and (\circ)0.24 mM.

SCh and calcium at the neuromuscular junction.

Materials and Methods

Ghost membranes were prepared from human erythrocytes according to the procedure of Hanahan⁸ with minor modifications. Fresh red blood cells were washed in 172 mM Tris buffer (pH 7.6) by centrifuging at 3,000 rpm for 5 min. The packed cells were hemolysed by forceful addition of 10 vol. of 11 mM Tris buffer (pH 7.6) under magnetically stirring and were followed twice by centrifugation at 10,000 rpm for 30 min at 4°C. Then the ghosts were spun down (20,000 rpm, 20 min, 4°C) and resuspended in 11 mM Tris buffer (pH 7.6). This procedure was repeated until ghost membranes became colorless. The final suspension was homogenized by a Potter-Elvehjem homogenizer (2.4 mg protein/ml), divided into 3 ml aliquots, and stored at -80°C until use. The negligible contamination of serum cholinesterase (called "pseudo-type" cholinesterase) in the ghosts was spectrophotometrically verified by the preliminary experiments with a pH indicator, bromothymol blue. Enzyme activity was

assayed by the method of Ellman et al.⁹ with acetylthiocholine as a substrate. The reaction mixture was composed of 5 mM 5,5' dithiobis-(2 nitrobenzoic acid), 0.2% Triton X-100 and 100 mM inorganic phosphate in a final volume of 2 ml (pH 7.4). Instead of phosphate buffer, 12 mM Tris was used in the experiment with calcium. The reaction was initiated by adding the substrate and the increase in absorbance at 412 nm was recorded by spectrophotometer, HITACHI model 320. All kinetic measurements, at least in duplicate, were carried out at 23°C. Protein concentration was determined by the method of Lowry et al.¹⁰ using BSA as a standard protein. The enzyme showed a K_m value of 7.1×10^{-5} M for acetylthiocholine.

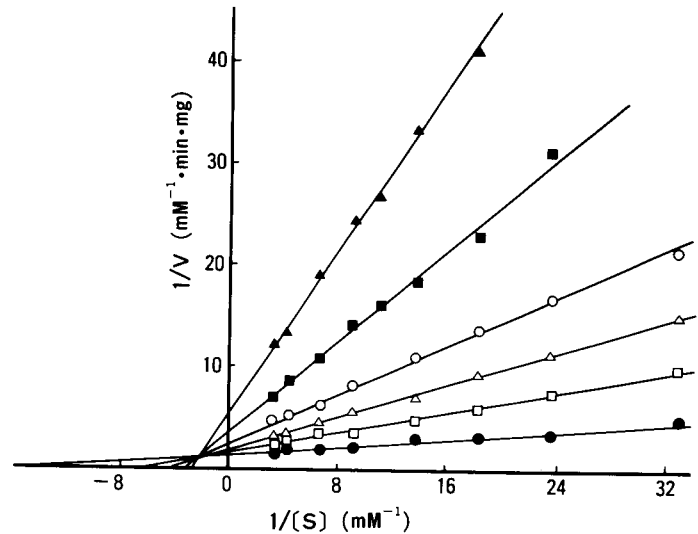
Chemicals were obtained as follows: acetylthiocholine chloride from Nakarai Chemical Co. Ltd., succinylcholine chloride from Kyorin Chemical Co. Ltd. and calcium chloride from Koso Chemical Co. Ltd. All other reagents were of analytical grade commercially available. SCh was dissolved in the reaction mixture immediately before use.

Results

Effect of SCh on AChE activity

SCh inhibited the AChE activity dose-dependently at low substrate concentrations (fig. 1). Inset shows the inhibition plot of AChE with SCh at 0.30 mM substrate. Half-maximal inactivation was observed at about 0.15 mM SCh. The K_i value from the Dixon plot was 6.7×10^{-5} M (data not shown). These values agree with those observed by Lanks and Sklar⁵ and by Foldes¹¹. The inhibition kinetics was investigated by double reciprocal plots of AChE activity at various concentrations of SCh (fig. 2). No intersection fell on one of the axes, showing a mixed type of inhibition by SCh. SCh possesses two trimethylammonium groups like decamethonium. One of the groups could bind to the catalytic site and the other to an allosteric site, inducing conformational changes in the active center. This may explain one of the kinetics of the SCh-AChE interaction. A clinical dose of SCh, according to Foldes¹¹, can achieve the plasma concentration of 10^{-5} M order. Therefore, our result would suggest that SCh influences tissue AChE, possibly

Fig. 2. Double-reciprocal plots of succinylcholine concentrations versus the rate of acetylthiocholine hydrolysis. Succinylcholine concentrations were (●) 0 mM, (□) 0.067 mM, (△) 0.13 mM, (○) 0.27 mM, (■) 0.53 mM and (▲) 1.07 mM.



by an allosteric mechanism, in the same concentration range as it affects cholinergic response, e.g., muscarinic effects.

Effect of calcium on AChE activity

Calcium suppressed the AChE activity dose-dependently (fig. 3). Inset shows the inhibition plot of AChE with calcium at 0.30 mM substrate. ID_{50} of calcium was 7.1 mM. The inhibition kinetics was investigated by the Lineweaver-Burk plotting of the enzyme activity with calcium (fig. 4). Calcium also exhibited a mixed type of inhibition. Dixon plot showed a biphasic curve with a bend at about 3 mM calcium and yielded two apparent K_i (fig. 5). In 12 mM Tris buffer (pH 7.4), K_i (1) was 1.2 mM and K_i (2) 3.1 mM. The data were highly reproducible and independent of order in mixing. Trace amounts of calcium ion in the assay medium would not contribute to the effects observed, since no alteration in the enzyme activity was obtained by adding EDTA (0.5 mM). According to Moore and Manery¹², Triton X-100 inhibited the membrane-bound AChE even in the low concentrations. In our experiment, however, the enzyme activity was not depressed by adding 0.2% Triton X-100 to ghost suspension before the assay. Then we performed the control experiment with and without Triton X-100 to confirm that the biphasic pattern was not caused by an interaction of calcium with this detergent. In the Dixon plot was again observed the

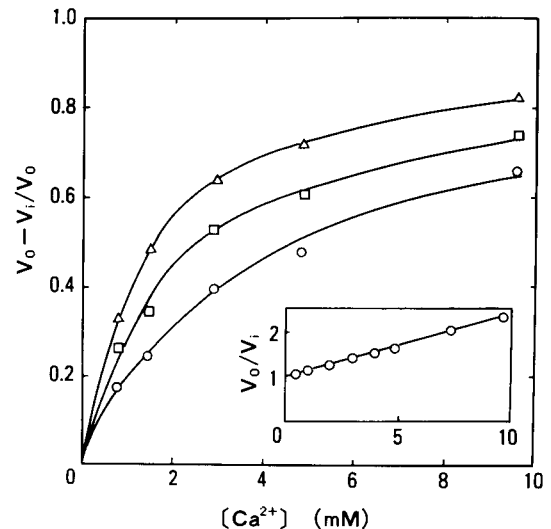


Fig. 3. Effect of calcium on hydrolysis of acetylthiocholine. Inset shows inhibition plot of acetylcholinesterase with calcium at 0.30 mM substrate. Half-maximal inactivation was observed at 7.1 mM calcium. V_i and V_0 represent the initial velocity with and without calcium, respectively. Substrate concentrations were (△) 0.030 mM, (□) 0.11 mM and (○) 0.24 mM.

biphasic inhibition pattern. As has been reported by Wermuth and Brodbeck¹³, this biphasic nature suggests two states of membrane-bound AChE. Applying the Hill equation to the biphasic pattern at 0.030 mM substrate, the coefficient

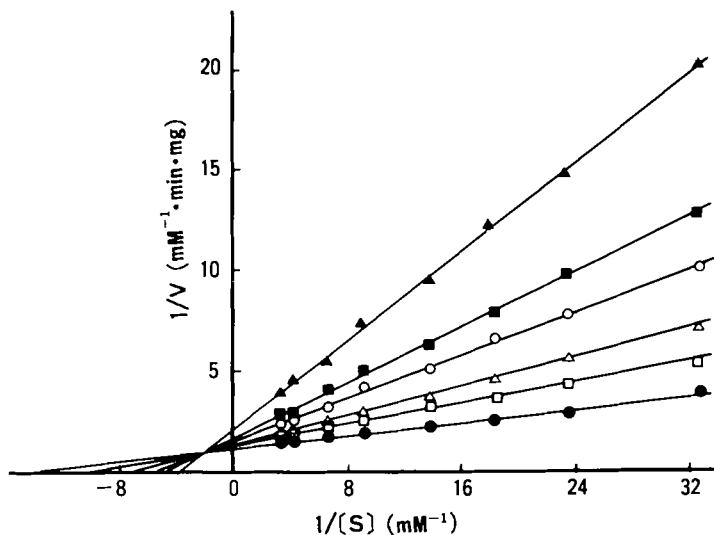


Fig. 4. Double-reciprocal plots of calcium concentrations versus the rate of acetylthiocholine hydrolysis. Calcium concentrations were (●)0mM, (◻) 0.7 mM, (△)1.4 mM, (○) 2.9 mM, (■)4.8 mM and (▲)9.6 mM.

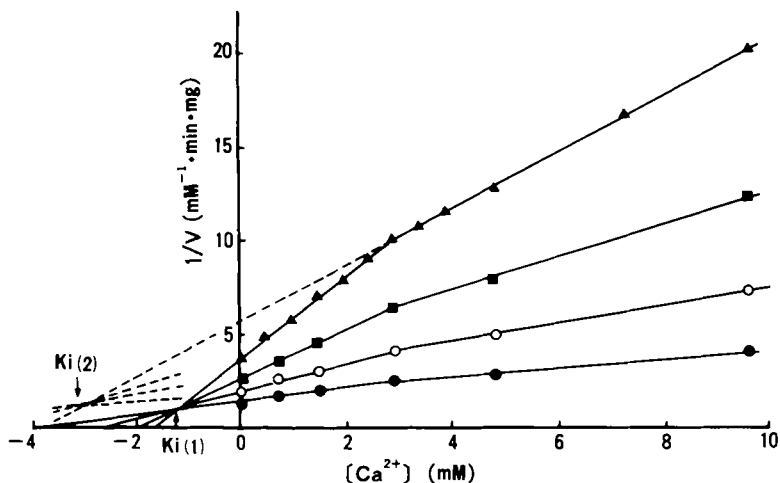


Fig. 5. Dixon plots of the effect of calcium on acetylthiocholine hydrolysis. Substrate concentrations were (▲)0.030 mM, and (■)0.54 mM, (○)0.11 mM and (●)0.30 mM.

would be 1.04 (state I) at the calcium concentration between 0 and 3.0 mM and 0.75 (state II) between 3.0 and 9.6 mM. However, based on the fact that physiological calcium concentration is lower than 2 mM, physiological significance of state II is in doubt.

Combined effects of calcium and SCh on AChE activity

In the presence of calcium, SCh inhibited the AChE activity, showing a hyperbolic curve in the plots (fig. 6). This may imply that SCh in combination with calcium exhibited a partially competitive kinetic for membrane-bound AChE at high SCh concentrations. At the higher calcium

concentrations, the inhibition curves shifted toward higher SCh concentrations range, indicating that calcium could reverse the inhibitory effect of SCh. Calcium increased greatly the enzyme activity in the presence of SCh (fig. 7). At the higher SCh concentrations, the greater reversing activity of calcium was observed. Referring to the clinical dosage, 1.5 mM calcium could increase the enzyme activity by approximately 25% at 6.7×10^{-5} M SCh. At calcium concentrations higher than 6 to 7 mM, however, the reversing activity was decreased.

Discussion

In this study, calcium exerted an inhibitory effect on membrane-bound AChE of human erythrocyte, unlike the previous observations^{6,14}, in which calcium served as an activator. effect on membrane-bound AChE of human erythrocyte, unlike the previous observations^{6, 14}, in which calcium serves as an activator. Their enzyme preparations are soluble form or mixed form of soluble and membrane-bound AChE. In general, AChE is membrane-associated and can hardly be solubilized. A review of literatures^{7,15,16} has indicated that preparation and purification procedures may alter the physical properties of membrane-bound enzymes and that their catalytic functions could be restored by addition of lipids. Also, Morero et al.¹⁷ have suggested that the membrane structure is an important factor for the allosteric changes of the membrane-bound enzymes. Thus, the discrepancy in the results may be explained by the different assay conditions. Ribaie and Kato¹⁸ also have studied the effects of calcium on soluble and membrane-bound AChE in *Electrophorus electricus*, and they have found that calcium enhances the hydrolysis rate of acetylcholine by soluble AChE and decreases it by membrane-bound AChE.

When the enzyme was exposed to SCh, calcium reversed the inhibitory effect of SCh on membrane-bound AChE. Some reports have shown that calcium antagonizes the effects of other muscle relaxants on AChE activity^{19,20}. Roufogalis, and Quist¹⁹ have reported that calcium reverses the inhibitory effect of decamethonium on partially purified AChE, whereas this reversal by calcium was not observed by Wins et al.²¹ in thier experiment using membrane-bound AChE from the same enzyme source as that of Roufogalis and Quist. Each subunit of AChE molecule is assumed to possess minimum five anionic subsites. In the terminology of Rosenberry²², one of them (site C) is catalytic anionic site and the others (site $P_1 - P_4$) are peripheral sites. Roufogalis and Quist¹⁹ have shown that calcium antagonizes decamethonium binding to soluble AChE in a competitive manner. Tomlinson et al.⁶ have suggested that decamethonium spans the C and P_1 site but calcium selectively binds to the

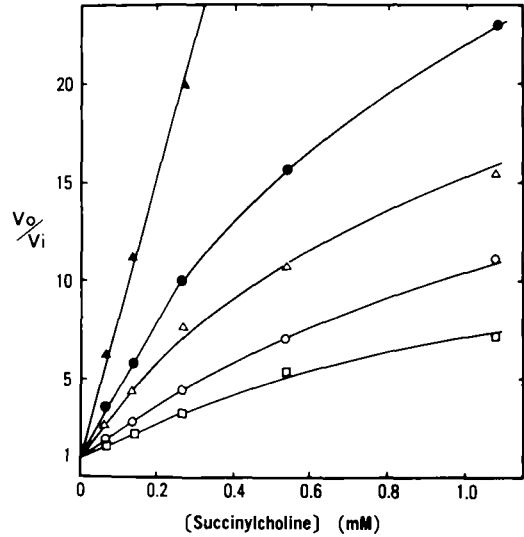


Fig. 6. Effect of succinylcholine on acetylcholinesterase activity in the presence of calcium. Calcium concentrations were (\blacktriangle) 0 mM, (\bullet) 1.4 mM, (\triangle) 2.8 mM, (\circ) 5.0 mM and (\square) 9.6 mM. Substrate concentration was 0.30 mM. V_i and V_0 represent the initial velocity with and without succinylcholine, respectively.

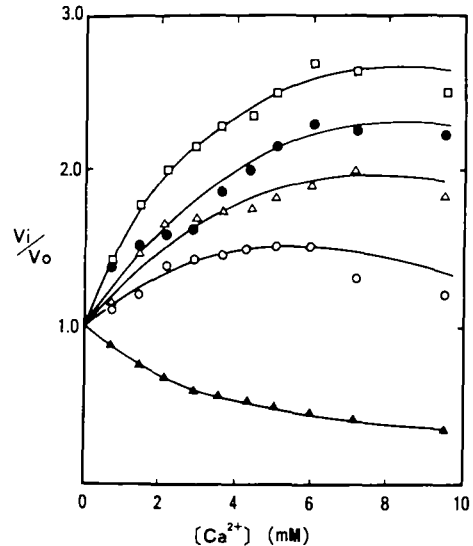


Fig. 7. Effect of calcium on the succinylcholine-induced depression of enzyme activity. Succinylcholine concentrations were (\blacktriangle) 0 mM, (\circ) 0.067 mM, (\triangle) 0.13 mM, (\bullet) 0.27 mM and (\square) 0.54 mM. Substrate concentration was 0.30 mM. V_i and V_0 represent the initial velocity with and without calcium, respectively.

P_1 site. Thus, P_1 site seems to function as an accelerator. In our data, however, calcium inhibited the activity of membrane-bound AChE. According to Robaire and Kato²³, moreover, membrane milieu did not significantly affect the enzymatic relationship between the peripheral site(s) and catalytic site of AChE in their study using soluble and membrane-bound enzymes. Therefore, several explanations are available for our data. The first is that calcium, only when together with SCh, would readily bind to the P_1 site. An alternate possibility is that the binding of calcium to allosteric sites except P_1 site would alter the conformation of the active center, by which SCh might lose the ability to bind to the C site and some allosteric sites.

At the neuromuscular junction, calcium enhances the release of acetylcholine from the motor nerve terminal and also stabilizes the postjunctional membranes³. This may explain why calcium antagonizes the neuromuscular blockade induced by magnesium or some antibiotics, especially in association with non-depolarizing muscle relaxants. However, SCh, depolarizing muscle relaxant, lacks an effective pharmacologic antagonist. In experimental animals, calcium antagonized the depolarizing block of SCh and potentiated the desensitization block, with its molecular mechanism being unknown^{2,3}. The latter block, or phase II block appears to be partially reversed by cholinesterase inhibitor. The pharmacological mechanism of phase II block has not been established yet, but several facts seem to be involved²⁴. One of them is that calcium influx through the opened ion channels can modify the physiological function of subendplate elements in muscles and of the acetylcholine receptors themselves²⁵. In our data calcium reversed the SCh-induced inhibition of AChE activity. Therefore, calcium will not be indicated for the treatment of phase II block. On the other hand, erythrocyte AChE itself seems to have an important role in removing the SCh administered intravenously²⁶. Erythrocyte enzyme will form a SCh-AChE complex and leave less available to act at the neuromuscular junction. If so, the antagonism by calcium of SCh-induced neuromuscular blockade^{1,2} may be partially

explained by our present results, in which calcium reversed the inhibitory effect of SCh on erythrocyte AChE activity.

Administration of SCh is accompanied by clinically significant adverse reactions. Recently, Nigrovic²⁶ has interpreted adverse hemodynamic reactions to SCh by an interaction between SCh and the presynaptic cholinceptors (nicotinic and muscarinic receptors) on the sympathetic nerve terminals. In his hypothesis, SCh usually produces balanced and clinical inconspicuous cardiovascular effects, because of the mutually opposing effects resulting from the activation of these cholinceptors by SCh. No sufficient evidence was presented to know whether the use of calcium could keep or lose this equilibrium between nicotinic and muscarinic effects, though AChE also has a high special activity in the cholinergic system and calcium enhances the release of acetylcholine presynaptically. On the other hand, SCh-induced postoperative myalgia, usually associated with an increase in serum potassium concentration and a decrease in calcium, was prevented by calcium pretreatment⁴. Further studies must be performed to elucidate this mechanism.

In conclusion, calcium reversed the inhibitory effect of SCh on erythrocyte AChE activity. It was suggested that the use of calcium may not be indicated for the reversal of SCh phase II block, while the exact physiological and clinical significance of these observations remains to be elucidated.

(Received Nov. 28, 1986, accepted for publication Nov. 28, 1986)

References

1. Mayrhofer OK: Prolonged apnea following scoline. *Anaesthesia* 7:250-251, 1952
2. Irwin RL, Wells JB, Whitehead RW: Effect of calcium on the duration of apnea induced by succinylcholine. *Anesthesiology* 17:759-767, 1956
3. Badola RP, Chatterji S, Pandey K, Kumar S: Effects of calcium on neuromuscular block by suxamethonium in dogs. *Br J Anaesth* 43:1027-1035, 1971
4. Shrivastava OP, Chatterji S, Kachhawa S, Daga SR: Calcium gluconate pretreatment for prevention of succinylcholine-induced myalgia. *Anesth Analg* 62:59-62, 1983
5. Lanks KW, Sklar GS: Inhibition of human

- erythrocyte acetylcholinesterase by succinylcholine. *Res Commun Chem Pathol Pharmacol* 14: 269–275, 1976
6. Tomlinson G, Mutus B, McLennan I: Activation and inactivation of acetylcholinesterase by metal ions. *Can J Biochem* 59:728–735, 1981
 7. Ott P: Membrane acetylcholinesterase: Purification, molecular properties and interactions with amphiphilic environments. *Biochim Biophys Acta* 822:375–392, 1985
 8. Hanahan DJ, Ekholm JE: The preparation of red cell ghosts (membranes), *Methods in Enzymology*. Vol 31. Edited by Fleischer S, Packer L. London, Academic Press, 1974, pp 168–172
 9. Ellman GL, Courtney KD, Andres V, Featherstone RM: A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochem Pharmacol* 7:88–95, 1961
 10. Lowry OH, Rosebrough NJ, Farr AL, Randall RJ: Protein measurement with the folin phenol reagent. *J Biol Chem* 193:265–275, 1951
 11. Foldes FF: Distribution and biotransformation of succinylcholine. *Int Anesth Clin* 13:101–115, 1975
 12. Moore RB, Manery JF: Resealing to small solutes of white erythrocyte membranes after incubation with EDTA, Ca²⁺, salt, sucrose, and phospholipase C. *Arch Biochem Biophys* 211:179–191, 1981
 13. Wermuth B, Brodbeck U: Interaction of proflavine and acriflavine with acetylcholinesterase. *Eur J Biochem* 37:377–388, 1973
 14. Heller M, Hanahan DJ: Human erythrocyte membrane bound enzyme acetylcholinesterase. *Biochim Biophys Acta* 255:251–272, 1972
 15. Coleman R: Membrane-bound enzymes and membrane ultrastructure. *Biochim Biophys Acta* 300:1–30, 1973
 16. Vessey DA, Zakim D: Regulation of microsomal enzymes by phospholipids. *J Biol Chem* 246: 4649–4656, 1971
 17. Morero RD, Bloj B, Farias RN, Trucco RE: The allosteric transitions from membrane-bound enzymes : Behavior of erythrocyte acetylcholinesterase from fat deficient rats. *Biochim Biophys Acta* 282:157–165, 1972
 18. Robaire B, Kato G: Effects of Mg²⁺ and Ca²⁺ on soluble and membrane-bound acetylcholinesterase from *Electrophorus electricus*. *Biochem Pharmacol* 23:2476–2480, 1974
 19. Roufogalis BD, Quist EE: Relative binding sites of pharmacological active ligands on bovine erythrocyte acetylcholinesterase. *Mol Pharmacol* 8:41–49, 1972
 20. Foidart JM, Gridelet J: Effects of procaine and d-tubocurarine on the activity of membrane bound acetylcholinesterase. *Biochem Pharmacol* 23:725–733, 1974
 21. Wins P, Schoffeniels E, Foidart JM: Inhibition of membrane-bound acetylcholinesterase by d-tubocurarine and its reversal by bivalent cations. *Life Sci* 9:259–267, 1970
 22. Rosenberry TL: Acetylcholinesterase, *Advances in Enzymology*. Vol 43, Edited by Meister A. New York, John Wiley & Sons, 1975, pp 190–196
 23. Robaire B, Kato G: Effects of edrophonium, eserine, decamethonium, d-tubocurarine, and gallamine on the kinetics of membrane-bound and solubilized eel acetylcholinesterase. *Mol Pharmacol* 11:722–734, 1975
 24. Standaert FG: Basic physiology and pharmacology of the neuromuscular junction, *Anesthesia*. 2nd Ed. Vol 2. Edited by Miller RD. New York, Churchill Livingstone, 1986, pp 835–869
 25. Salpeter MM, Leonard JP, Kasprzak H: Agonist-induced postsynaptic myopathy. *Neurosci Commun* 1:73, 1982
 26. Lehmann H, Liddell J: The cholinesterases, *Modern Trends in Anesthesia*. Vol 2. Edited by Evans FT, Gray TC. London, Butterworth, 1962, pp 188–193
 27. Nigrovic V: Succinylcholine, cholinceptors and catecholamines : Proposed mechanism of early adverse haemodynamic reactions : *Can Anaesth Soc J* 31:382–394, 1984